

Summary

The influence of anaerobiose and of varied oxygen tensions upon respiration and histamine release during anaphylactic reaction in guineapig lung slices *in vitro* was studied.

No histamine was liberated in the absence of oxygen. With increasing oxygen tensions, increasing histamine quantities were released; these quantities reached the control values when the atmosphere contained 10% oxygen.

These results indicate that the mechanism of histamine release in the anaphylactic reaction is linked with the aerobic metabolism of the cell.

Effect

of Reserpine Analogues on Tissue Serotonin

In a recent note, it was reported that a differential effect was exerted on brain serotonin by reserpine and its analogues: carbethoxysyringoyl methyl reserpate (SU 3118 or syrosingopine) and methyl-18-O(3-N,N-dimethylaminobenzoyl) reserpate (SU 5171). Reserpine and SU 5171 release brain serotonin and induce sedation, while SU 3118, an hypotensive agent without sedative effect, is unable to release brain serotonin¹⁻³.

Since a poor penetration of SU 3118 into the brain might account for the lack of brain serotonin depletion, the effect of SU 3118, in comparison with SU 5171 and reserpine, on serotonin content of other organs has been investigated.

The method used was the following: Sprague-Dawley rats of the average weight of 200 g were injected intraperitoneally with different concentrations of reserpine (kindly supplied by Dr. V. Ghetti, CIBA, S.p.A., Milano) or SU 3118, or SU 5171 (received through the courtesy of Dr. A. J. PLUMMER, CIBA Labs., Summit, N. J.). 4 h after the administration, the animals were killed by decapitation and the serotonin content of brain, intestine, kidney, lung, and spleen was determined by the method of BOGDANSKI *et al.*⁴ employing a Farrand spectrophotofluorimeter.

The results obtained are reported in a Table.

¹ R. A. MAXWELL, H. POVALSKI, and A. J. PLUMMER, *J. Pharmacol. exper. Therap.* **125**, 178 (1959).

² K. F. FINGER, F. B. HUGHES, and B. B. BRODIE, *Fed. Proc.* **18**, 388 (1959).

³ S. GARATTINI, A. MORTARI, A. VALSECCHI, and L. VALZELLI, *Nature* **183**, 1273 (1959).

⁴ D. F. BOGDANSKI, A. PLETSCHE, B. B. BRODIE and S. UDEN-FRIEND, *J. Pharmacol. exper. Therap.* **117**, 82 (1956).

While reserpine and SU 5171 deplete serotonin in all the organs tested (except kidney in the case of SU 5171) SU 3118, at a concentration of 20 mg/kg lowers only lung and spleen serotonin. 20 mg/kg of SU3118 are less effective than 5 mg/kg of reserpine in releasing intestinal serotonin.

These results suggest that lung and spleen are more sensitive than other organs to the action of SU 3118. Under the action of SU 3118, the cerebral, intestinal, and renal serotonin behave similarly. Therefore, the hypothesis of a resistance of the blood-brain barrier to SU 3118 seems unlikely.

S. GARATTINI, R. KATO, and L. VALZELLI

Istituto di Farmacologia e di Terapia, Università degli Studi, Via A. Del Sarto 21, Milano, July 11, 1959.

Riassunto

La differenza di comportamento dimostrata, nel ridurre il tasso di serotonina di diversi organi, da due derivati reserpinici (SU 3118–SU 5171) non sembra potersi ricondurre solo ad una differente capacità di attraversamento della barriera ematoencefalica da parte delle due molecole in istudio.

The Stereochemistry of some Naturally Occurring Guaianolides and Isophotosantonin Lactone

Recent work on the stereochemistry of cycloheptanes^{1,2} has pointed out the general similarity of these systems to the more widely studied cyclohexane compounds. The application of the principles discussed by these workers, namely that a substituent group in a cycloheptane prefers the *quasi*-equatorial conformation, and that elimination reactions proceed *via* the same *trans-anti*-parallel route as in cyclohexanes, coupled with the logical extension from the cyclohexane field that a *cis* fused γ -lactone is more stable than the corresponding *trans* isomer, permits the deduction of provisional stereochemical relationships in several sesquiterpenic lactones. BARTON³ has already assigned structure I to geigerin on the basis of rotatory dispersion data⁴, and an assumption regarding the nature of the lactone ring similar to that which we have made.

¹ J. W. HUFFMAN and J. E. ENGLE, *J. org. Chem.*, in press.

² N. L. ALLINGER, *J. Amer. chem. Soc.* **81**, 231 (1959).

³ D. H. R. BARTON and J. E. D. LEVISALLES, *J. chem. Soc.*, **1958**, 4518.

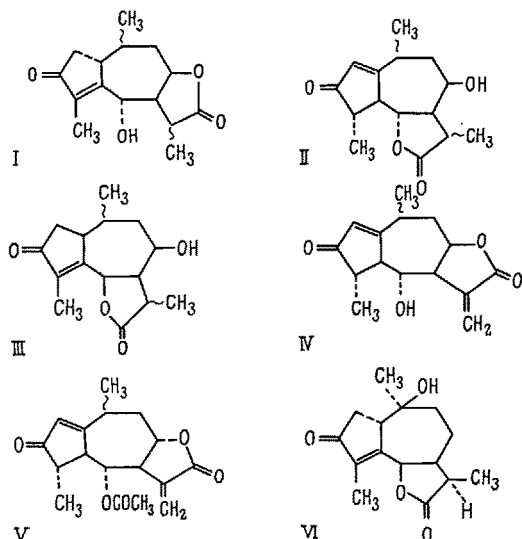
⁴ C. DJERASSI, J. OSIECKI, and W. HERZ, *J. org. chem.* **22**, 1361 (1957).

Treatment	Dose mg/kg i. p.	Serotonin content (μ /g \pm S. E.)				
		Brain	Intestine	Kidney	Lung	Spleen
Controls	—	0.43 \pm 0.08	4.76 \pm 0.24	0.34 \pm 0.03	2.86 \pm 0.16	3.51 \pm 0.24
Reserpine	2.5	0.17 \pm 0.005	2.95 \pm 0.15	0.19 \pm 0.02	1.6 \pm 0.23	1.09 \pm 0.24
Reserpine	5	0.12 \pm 0.006	2.04 \pm 0.12	—	—	—
SU 3118	20	0.46 \pm 0.005	3.37 \pm 0.36	0.29 \pm 0.03	1.64 \pm 0.27	1.04 \pm 0.09
SU 3118	50	0.13 \pm 0.011	2.64 \pm 0.05	—	—	—
SU 5171	5	0.25 \pm 0.009	3.98 \pm 0.24	0.49 \pm 0.07	1.84 \pm 0.28	2.23 \pm 0.73
SU 5171	7.5	0.12 \pm 0.005	2.73 \pm 0.21	0.28 \pm 0.02	1.21 \pm 0.07	0.59 \pm 0.03

Each value represents the average of at least 8 determinations.

The conformation of I is fixed by the ring fusion, and demands that the carbon-carbon bond at C₁ be *quasi-equatorial* to the seven-membered ring. Consequently the substituents at C₆, C₇, and C₈ will be *e'*, *e'*, and *a'*, respectively.

The assignment of stereochemistry to the remaining substances, desacetylisenotenulin (II), helenalin (IV), balduilin (V, or the equivalent structure with the lactone and acetoxyl interchanged), and isophotosantonin lactone (VI), rests on the following arguments. All of these substances have been shown to have the configuration indicated at C₁, C₄ and C₅ by rotatory dispersion studies⁴. In addition tenulin, helenalin, and balduilin have the same configuration at C₇ and C₁₀⁵. These compounds, as well as geigerin (I), and isophotosantonin lactone (VI) are all readily hydrogenated to saturated ketones under mild conditions^{3,4}. On the other hand, desacetylisenotenulin (III) which has unnatural stereochemistry at C₁ and is formed under conditions which should not disturb the configuration at C₇, is not readily hydrogenated^{4,6}. During this interconversion, in order to maintain the *e'* conformation at the ring fusion it is necessary for the seven membered ring to flip to an alternate conformation, causing all the other substituent groups to undergo corresponding conformational changes. The differences in ease of hydrogenation may then be explained by the presence of a bulky *e'* group at C₇ in tenulin, being changed to *a'* in desacetylisenotenulin. This group then shields the double bond from hydrogenation. On this basis the stereochemistry at C₇ is assigned as shown in desacetylisenotenulin, helenalin, balduilin, and isophotosantonin lactone.



The assignment of configuration to the hydroxyl groups rests upon the following observations. Dihydroisotenulin on basic hydrolysis and subsequent reacidification affords desacetyldihydroisotenulin and an isomeric lactone resulting from lactonization at C₈. On this basis the oxygens at C₆ and C₈ must be equivalent (*cis*), and on the basis of our assumptions regarding the stereochemistry of these lactones, *vide supra*, they must also be *cis* to the carbon chain at C₇. This gives rise to the partial stereochemistry for desacetylisenotenulin as illustrated (II).

⁵ W. HERZ, R. B. MITRA, and P. JAYARAMAN, Abstr. Papers 136th National Meeting of the American Chemical Society, Sept. 13-18 (1959), p. 50 P.

⁶ B. H. BRAUN, W. HERZ, and K. RABINDRAN, J. Amer. chem. Soc. 78, 4423 (1956).

Helenalin (IV), and its derivatives, on hydrolysis and reacidification are recovered unchanged^{7,8} indicating that the groups at C₇ and C₈ are *cis*, and that the C₆ hydroxyl is probably *trans* to the side chain. HERZ has recently shown that balduilin (V) is epimeric with tenulin at C₈, and helenalin at C₈⁵, giving balduilin the structure shown.

The stereochemistry at C₁₀ in isophotosantonin lactone (VI) rests upon the following argument. Dehydration of (VI) with thionyl chloride in pyridine affords an exocyclic, non-conjugated olefin⁸, while acid catalyzed dehydration gives the doubly unsaturated, conjugated ketone. On this basis we propose that the C₁₀ hydroxyl group is *cis* to the hydrogen at C₁, otherwise dehydration with thionyl chloride should have afforded the conjugated isomer *via* a *trans* elimination involving the activated hydrogen at C₁. The configuration at C₈ is assigned as shown, with inversion taking place during the formation of VI from santonin by analogy with the santonin-desmotroposantonin conversion⁹. The stereochemistry of the methyl group in the side chain is considered to be unchanged from that in santonin¹⁰, in view of the fact that the conditions for the formation of VI from santonin do not appear vigorous enough to cause epimerization of a group adjacent to a saturated carbonyl. A similar argument may be used in support of the proposed configuration at C₇, which is the same as that in santonin.

J. W. HUFFMAN

School of Chemistry, Georgia Institute of Technology, Atlanta (Georgia), August 17, 1959.

Zusammenfassung

Die Stereochemie der Sesquiterpene Tenulin, Balduilin und Helenalin sowie isophotosantonisches Lakton wird behandelt und provisorische Strukturformeln für diese Substanzen vorgeschlagen.

⁷ R. ADAMS and W. HERZ, J. Amer. chem. Soc. 71, 2546 (1949).

⁸ D. H. R. BARTON, P. de MAYO, and M. SHAFIQ, J. chem. Soc. 1957, 929.

⁹ HUANG-MINLON, J. Amer. chem. Soc. 70, 611 (1948).

¹⁰ R. B. WOODWARD and P. YATES, Chem. & Ind. 1954, 1319. — E. J. COREY, J. Amer. chem. Soc. 77, 1044 (1955).

The Effect of Insulin on Production of Granulation Tissue in Rats¹

The effect of hormones on connective tissue has been extensively studied. The formation of granulation tissue — one of the basic reactions of the mesenchyme — is influenced by several hormones. The stimulation of this process by STH, thyroxine, and aldosterone and the inhibition by cortisone and sexual steroids is well known²⁻⁹.

¹ Presented at the 25th meeting of the Hungarian Physiological Society, Szeged, July 1959.

² M. TAUBENHAUS and G. D. AMROMIN, J. Lab. clin. Med. 36, 7 (1950).

³ M. TAUBENHAUS and G. D. AMROMIN, Endocrinol. 44, 359 (1949).

⁴ P. DESAULLES, Z. ges. exp. Med. 124, 30 (1954).

⁵ P. DESAULLES, W. SCHULER, and R. MEIER, Exper. 11, 68 (1955).

⁶ T. H. RINDANI, Arch. int. Pharmacodyn. 99, 467 (1954).

⁷ C. RAGAN, E. L. HOWES, C. M. PLOTZ, K. MEYER, and J. W. BLUNT, Proc. Soc. exp. Biol. Med., N. Y. 72, 718 (1949).

⁸ G. DOMBRÁDI and S. KARÁDY, Bőrgyógy. Venerol. Szeml. 3, 79 (1953).

⁹ G. DOMBRÁDI and S. KARÁDY, Acta physiol. Acad. hung., Suppl. 5, 92 (1954).